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ORIGINAL ARTICLE

Influence of Azolla (*Azolla microphylla* Kaulf.) compost on biogenic gas production, inorganic nitrogen and growth of upland kangkong (*Ipomoea aquatica* Forsk.) in a silt loam soil

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Abstract

Azolla microphylla Kaulf. (Azolla) biomass was composted to create a high nitrogen (N) organic matter amendment (Azolla compost). We examined the effect of this Azolla compost on carbon (C) and N mineralization and the production of biogenic gases, nitrous oxide (N₂O) and carbon dioxide (CO₂), in a soil incubation experiment. A pot experiment with upland kangkong (*Ipomoea aquatica* Forsk.) examined plant growth in silt loam soil treated with three levels of Azolla compost. The results showed that N₂O production from soil increased with urea amendment, but not with Azolla compost treatments. The Azollaamended soil showed enhanced CO₂ production throughout the 4-week incubation. The Azolla-treated soils showed a 98% lower global warming potential compared to urea treatment over the 4-week incubation. However, Azolla-amended soil had higher nitrate (NO₃⁻) levels compared to urea-fertilized soil at 1 week of incubation, and these were maintained until the fourth week. Soils amended with Azolla compost showed lower ammonium nitrogen (NH₄-N) levels than those in the urea-fertilized soils. The height and dry weight of upland kangkong fertilized with Azolla compost were similar to plants receiving urea fertilization. Therefore, the use of Azolla compost as a substitute for urea fertilizer would be beneficial for reducing the production of N₂O while maintaining plant growth.

Key words: Azolla, compost, greenhouse gas, kangkong, nitrous oxide.

INTRODUCTION

Applications of organic amendments to soils are useful as alternatives to chemical fertilizers for increasing crop productivity as well as improving long-term soil fertility and quality. These organic amendments may be crucial sources of fertility for small landholders who comprise the majority of farmers worldwide, and yet may lack resources such as mineral fertilizers (Herrero *et al.* 2010) or prefer organic management for environmental residues provides a source of readily available carbon (C) and nitrogen (N) in the soil, and subsequently influences carbon dioxide (CO₂) and nitrous oxide (N₂O) production (Padre *et al.* 2005; Lou *et al.* 2007; Toma and Hatano 2007; Kimura *et al.* 2011; Zhu *et al.* 2013). Several factors have been identified that affect the rate of N₂O emission from agricultural systems, including N amendment (Bouwman 1996; Jumadi *et al.* 2008a, 2008b; Fukumoto and Inubushi 2009), pH (Xu and Inubushi 2005), temperature (Lang *et al.* 2011), organic amendment (Lou *et al.* 2007; Vano *et al.* 2011, Thangarajan *et al.* 2013), soil moisture (Kirk *et al.* 2013; Zhu *et al.* 2013) and land use management (Jumadi *et al.* 2005; Lang *et al.* 2011; Kong *et al.* 2013). Organic amendments or fertilizers affect N₂O gas

reasons. Incorporation of organic amendments or crop

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production in several ways: (1) the type of N [nitrate (NO_3^-) , ammonium (NH_4^+) or organic N] affects N gas production during nitrification and denitrification; (2) the presence of easily decomposable organic C enhances soil respiration and biological oxygen (O_2) demand inducing low-O₂ environments and subsequent denitrification; (3) the addition of other compounds (such as salt and water) have effects on biological, chemical and physical soil processes (Velthof *et al.* 2012; Thangarajan *et al.* 2013).

Azolla (Azolla microphylla Kaulf.), a small floating water fern, is commonly seen in lowland rice fields and has a symbiotic association with nitrogen-fixing Cyanobacteria (i.e. Anabaena azollae), which inhabit the dorsal lobe of the leaves. The benefit of including Azolla in agricultural systems is primarily due to its high N fixation capacity with amounts up to 1.2 kg N ha⁻¹ daily (Talley et al. 1977). Azolla is widely used as a biofertilizer for rice in China, Vietnam, Indonesia, Thailand and other East and South Asian countries (Cheng et al. 2010), and has also been used as a source of green manure and compost (Bordoloi et al. 2007). Although the growth and use of Azolla in South Sulawesi Province, Indonesia, has not been surveyed, based on our observations, Azolla is widespread in ponds and standing water and spreads rapidly in rice fields. Furthermore, Djojosuwito (2000) has reported that when approximately 500 kg of Azolla seed was spread in 1 ha of a rice field, the Azolla increased up to 20,000 kg ha⁻¹ within 2 weeks. This suggests that Azolla has the capacity to quickly more than double, or increase up to 40-fold, the biomass of the initial seed. Composting is the controlled microbial aerobic decomposition and stabilization of organic substrates, under conditions that allow the generation of high temperatures by thermophilic microbes, to obtain an end product that is stable, free of pathogens and viable weed seeds, and suitable for crops. These biological processes also change the composition of the organic matter, particularly reducing its content of soluble organic compounds and its C to N ratio. Composting plant residues therefore affects N₂O, CO₂ and nitric oxide (NO) emissions processes compared to direct application of the same residues (Velthof et al. 2012; Kirk et al. 2013; Zhu et al. 2013).

Upland kangkong (*Ipomoea aquatica* Forsk.) is widely cultivated in Indonesia and has become a popular leafy vegetable compared to water spinach (*Ipomoea aquatica* Forsk.). Recently, there has been an increased demand for organic upland kangkong due to perceived health benefits. Upland kangkong often requires supplemental N for improved quality of growth. Organic amendments such as compost are important options for maintaining soil fertility and N supply.

The application of Azolla compost to soils has been proposed as a useful source of N for crops and as an amendment for increasing the organic C content of soils with low organic matter (Lumpkin and Plucknett 1980; Azmal *et al.* 1996, 1997). Some compost with high C to N ratios may promote N immobilization in soils (Norton and Schimel 2011; Zhu *et al.* 2013) and it may be appropriate to add synthetic N fertilizer to these composts in order to increase the availability of N to crops. There is, however, still little available information concerning the impact of Azolla compost applications on N availability and greenhouse gas emissions from soils. Laboratory incubation experiments were performed to understand the influence of Azolla compost on soil N levels and N₂O and CO₂ emissions. Plant growth experiments were conducted to examine the effects of the Azolla compost on the growth of upland kangkong.

MATERIALS AND METHODS

Soil analysis

The soil was sampled at a vegetable farm in Gowa district, South-Sulawesi Province, Indonesia, in 2011, and would be classified in the Entisol soil order (Soil Survey Staff 1999). Soil samples were taken in triplicate at a depth of 0-20 cm, passed through a 2-mm mesh sieve, and stored at 4°C until used. Soil and compost analyses were performed at the Soil Testing Laboratory (Department of Agriculture, South Sulawesi Province, Indonesia). Soil textures were determined by hydrometer method (Bouyoucos 1962). Soil pH was determined in a 1:5 (soil: 1M potassium chloride, KCl) using a glass electrode. Total organic N and organic C were analyzed by Kjeldahl and combustion methods, respectively (Page et al. 1982). Cation exchange capacity (CEC) was determined according to Burt (2004), and extractable phosphorus (phosphorus pentoxide, P₂O₅) by the Bray method (Bray and Kurtz 1945). Ammonium nitrogen (NH_4^+-N) and nitrate nitrogen (NO_3^--N) were extracted with 2 M KCl and analyzed by colorimetric methods (Rayment and Higginson 1992). All physiochemical properties and gas production are expressed on a dry soil weight basis.

Preparation of Azolla compost

Two kilograms of Azolla (Azolla microphyla) plant material was collected on December 5, 2011, from a rice field in Maros district (5m above sea level, 05° 00.419' S 119° 31.219' E), South-Sulawesi Province, Indonesia. Azolla biomass was washed with distilled water three times, then dried in the oven at 55°C until the moisture content was around 50%. The dried Azolla biomass was placed in a black plastic bucket and 250 mL of molasses was added and mixed, then the bucket was covered with black plastic. The composting process continued for 1 week, and if the temperature of the material increased substantially the material was shifted into another bucket. The harvested Azolla compost was dried in the oven at 55°C, and then ground to pass a 2.00-mm sieve before analysis.

Soil incubation

A composited soil sample was preincubated at 27°C for 7 d and then kept at 55% of water holding capacity (WHC) and treated with the N source amendments. The treatments were: control (no N added), Azolla compost low rate (Azolla low) equivalent to 80.3 g N m⁻² rate or about 803 kg N ha⁻¹ (equivalent to urea 5 g N m⁻² or approximately 50 kg N ha⁻¹), Azolla compost medium rate (Azolla med) equivalent to 160.5 g N m⁻² rate or 1605 kg N ha⁻¹ (equivalent to urea 10 g N m⁻² or approximately 100 kg N ha⁻¹), Azolla compost high rate (Azolla high) equivalent to 240.8 g N m⁻² rate or 2408 kg N ha⁻¹ (equivalent to urea 20 g m⁻² or approximately 200 kg ha⁻¹) and urea equivalent to 10 g N m⁻² or approximately 60 µg N g⁻¹ soil. Fifteen grams of soil were incubated aerobically at 27°C in sealed 120-mL bottles for 28 days in triplicate. Production of N2O, CO₂ gases and inorganic N as ammonium and nitrate were determined for each soil sample. Every 7 d, the gases in the headspace of each bottle were taken and transferred to 22-mL vacuum vials; the incubation bottles were then aerated and resealed for later sampling. Five grams of soil was extracted for inorganic N with 1M KCl solution (1:5) and these extracts were analyzed for ammonium and nitrate colorimetrically as described above. Gas samples were sent to the Laboratory of Soil Science, Chiba University, Japan, to quantify the concentrations of N₂O and CO₂ using gas chromatographs (Shimadzu, GC 14B, Kyoto, Japan) equipped with an electron capture detector (ECD) and a thermal conductivity detector (TCD), respectively. The global warming potential (GWP) was calculated according to IPCC (2007) which was done as sum of N₂O production during incubation time and then converted to factor as 1 kg N₂O is equivalent to 298 kg CO₂ at 100 years' time horizon. The mitigation potential was assessed as the percentage of reduction in N₂O production from soils treated with urea compared to those treated with Azolla compost.

Upland Kangkong growth

Pot experiments were conducted in the field at the Biology Department Farm, Makassar State University, during January to March 2012. The randomized block experimental design consisted of four treatments plus one control (check), each with three replicates; the experiment was repeated twice (30 pots total). The polyvinyl chloride (PVC) pots had an inner diameter of 22.5 cm and a depth of 20.5 cm. The treatments and soil characteristics were the same as those used in the incubation experiment. Azolla compost and urea were applied at one time by spreading and incorporation to the surface soil 5 d after sowing seeds of upland kangkong. The growth of upland kangkong was determined after 35 d. Plant height (cm) was measured weekly and, at the end of the growth period, the aboveground plant material was harvested and the dry weight was determined (Singla et al. 2013). Statistical analysis was performed using SPSS version 16.0 (IBM Corp 2008). Posthoc comparisons were examined by least significant difference (LSD = 0.05).

RESULTS

Soil and Azolla compost characteristics

The soil was a silt loam with initial physicochemical properties as follows: texture; 26% sand, 47% silt, and 27% clay; pH (KCl 1:5) 4.1; total organic C 1.47%; total organic N 0.18% and C/N ratio 8.2; CEC 28.13 (cmol_c kg⁻¹); extractable phosphorus 50 mg P₂O₅ g⁻¹ soil; NH₄⁺-N 4.08 µg g⁻¹ dry soil; and NO₃⁻N 7.51 µg g⁻¹ dry soil. These characteristics are typical for upland soils under agriculture with the use of chemical fertilizers. The total organic C and N of Azolla compost were 27.7 and 2.7%, respectively, resulting in a C/N ratio of 10.

Production of N₂O and CO₂, global warming potential and inorganic N in a silt loam soil

The production of nitrous oxide (N₂O) gas was significantly increased by the addition of urea compared to the Azolla compost-amended or control soils. The Azolla amendment did not significantly affect the N₂O production during the 4-week incubation period (Fig. 1). N₂O production in all treatments peaked at the 3-week sampling, except for the high-level Azolla compost which was highest at the 1-week sampling and then decreased. The urea addition exhibited the highest production of N₂O for the third week of the incubation (890 ng g⁻¹), then decreased to 234 ng g⁻¹ in the fourth week.

The production of CO_2 from the treated soils is given in Fig. 2. Amendments of Azolla compost and urea remarkably enhanced CO_2 production. In the first week of incubation, the soil respiration was significant higher for the medium- and high-level Azolla compost treatments. During the third and fourth weeks, the Azolla high and urea-treated soils both showed elevated CO_2 Influence of Azolla compost on N₂O, CO₂ and plant growth 725

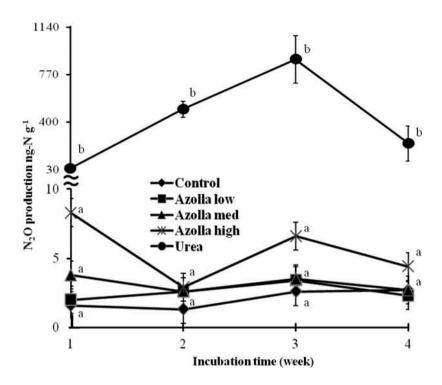


Figure 1 Production of nitrous oxide (N₂O) during soil incubations sampled weekly. Value followed by different letters indicate significant different (least significant difference, LSD; P < 0.05). The error bars indicate the standard deviations. N, nitrogen.

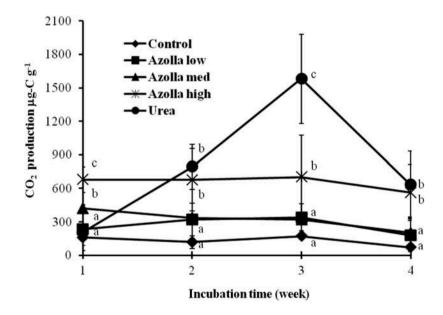


Figure 2 Production of carbon dioxide (CO₂) during soil incubations sampled weekly. Value followed by different letters indicate significant different (least significant difference, LSD; P < 0.05). The error bars indicate the standard deviations.

production. Soil treated with urea significantly contributed to GWP (966 kg CO_2 ha⁻¹) compared to other treatments: Azolla low with 6 kg CO_2 ha⁻¹, Azolla medium with 7 kg CO_2 ha⁻¹ and Azolla high with 13 kg CO_2 ha⁻¹. The results also showed that the reduction of sources to atmospheric radiative forcing of CO_2 gas was approximately 98–99%. Cumulative levels of inorganic N are shown in Tables 1 and 2. During the first week, the Azolla medium- and high-level amendments showed elevated nitrate levels. During the second and third

Table 1 Levels of ammonium (NH_4^+) (µg N g⁻¹) accumulated during incubation of the silt loam soil

	Concentration of NH4 ⁺ (µg N g ⁻¹) incubation time (week)			
Treatments	1	2	3	4
Control Azolla low Azolla med Azolla high Urea	$4.1^{a} \\ 4.5^{ab} \\ 5.1^{ab} \\ 6.4^{b} \\ 19.8^{c}$	$ \begin{array}{r} 1.8^{a} \\ 2.6^{a} \\ 2.7^{a} \\ 6.3^{b} \\ 15.2^{c} \end{array} $	$1.9^{a} \\ 2.5^{ab} \\ 3.5^{ab} \\ 4.1^{b} \\ 10.2^{c}$	$ \begin{array}{r} 1.5^{a} \\ 2.5^{a} \\ 2.6^{a} \\ 3.5^{a} \\ 6.1^{b} \end{array} $

N, nitrogen. Values followed by different letters in each column indicate significant differences (least significant difference, LSD; P < 0.05).

Table 2 Levels of nitrate (NO_3^-) ($\mu g \ N \ g^{-1})$ accumulated during incubation of the silt loam soil

	Concentration of NO_3^- (µg N g ⁻¹) incubation time (week)			
Treatments	1	2	3	4
Control Azolla low Azolla med Azolla high Urea	$\begin{array}{c} 46.9^{a} \\ 72.8^{ab} \\ 74.3^{b} \\ 82.8^{b} \\ 63.0^{ab} \end{array}$	$41.6^{a} \\ 89.4^{b} \\ 90.2^{b} \\ 98.6^{b} \\ 79.4^{b}$	57.1^{a} 84.3 ^b 88.5 ^b 116 ^c 166 ^d	$\begin{array}{r} 68.9^{a} \\ 126^{b} \\ 146^{b} \\ 158^{b} \\ 33.5^{a} \end{array}$

N, nitrogen. Values followed by different letters in each column are significantly different (least significant difference, LSD; P < 0.05).

weeks, all Azolla compost treatments and the urea treatment had elevated nitrate, which persisted for the Azolla treatments through the incubation period. By the fourth week, the urea treatment nitrate levels were not significantly different from those of the control soil. Soils amended with Azolla compost showed lower NH₄-N levels than those in the urea-fertilized soils.

Growth of upland kangkong

The height and dry weight of kakong plants revealed that both Azolla compost and urea can be used to increase the plant growth of upland Kakong as compared to growth in non-amended soils (Fig. 3 and Table 3). Azolla compost treatments enhanced the height of plants starting in the third week after sowing and it remained significantly higher through harvest. At the end of the growth period, the dry weight of the plants receiving Azolla compost was not significantly different than the control or the urea treatment, while the urea-fertilized plants were significantly higher than the control.

DISCUSSION

The Azolla material rapidly composted due to thin leaves and low recalcitrance. Several studies reported that the C/N values of mature composts are between 10 and 15 (Rynk 1992). Zbytniewski and Buszewski (2005)

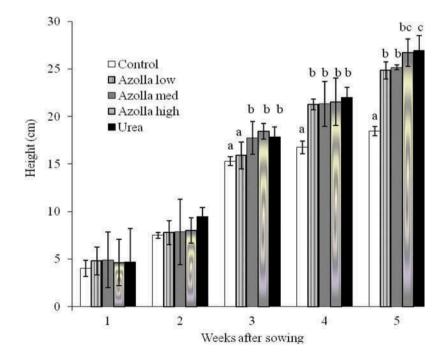


Figure 3 Height (cm) of upland kangkong (*Ipomoea aquatica* Forsk.) grown in pots. Treatments are: control (no amendment), Azolla Compost low (80.3 g m⁻²), Azolla Compost medium (160.5 g m⁻²), Azolla Compost high (240.8 g m⁻²), Urea (10 g N m⁻²). Different letters refer to significant differences (least significant difference, LSD; P < 0.05).

Table 3 Plant biomass (dry weight, grams per pot) of aboveground upland kangkong (*Ipomoea aquatica* Forsk.) studied in a pot experiment. Soil treatments are the same as for Fig. 1. Different letters refer to significant differences (least significant difference, LSD; P < 0.05)

Treatments	Plant biomass (dry weight) (gram pot ⁻¹)
Control Azolla low Azolla med Azolla high Urea	$\begin{array}{c} 4.77^{a} \\ 5.13^{ab} \\ 5.30^{ab} \\ 5.38^{ab} \\ 5.42^{b} \end{array}$

Soil treatments are the same as for Fig. 3. Values followed by different letters in each column are significantly different (LSD, P < 0.05).

reported that a C/N ratio of about 15 expresses stabilization of composting mass, and one below 12 indicates a high degree of compost maturity. In addition, Komilis and Tziouvaras (2009) and Komilis *et al.* (2011) also suggested that a C/N ratio of 10 indicates mature composts. Hence with a C/N ratio of 10, we judged the maturity and stability of the *Azolla* compost as suitable for our research purposes.

The high CO₂ and N₂O release from the urea treatment may indicate the loss of N through nitrification or denitrification processes in the urea treated soils. The amendment of organic matter can substitute for or supplement mineral N fertilizer in agricultural systems; however, the management of the timing of nutrient release remains a challenge. Soil organic matter (SOM) content and texture are important factors affecting C and N mineralization under constant soil moisture (Zhu *et al.* 2013). The N and C available to soil from an amendment are influenced by a complex interaction of amendment characteristics, soil characteristics and the rate of application (Zhu *et al.* 2013).

The Azolla compost used was produced using the addition of a readily decomposable soluble organic C (molasses) which promoted the immobilization of the available N into microbial biomass and products. Therefore, the Azolla compost used with a relatively low C/N of 10 should release N relatively rapidly into the soil. This addition of readily decomposable organic matter increased microbial activity as indicated by CO₂ production. Compost addition typically leads to the buildup of soil organic matter and the formation of macro and micro aggregates (Shibahara and Inubushi 1997; Zaman *et al.* 2002).

The C/N ratio of amendments may also be useful in predicting N mineralization (Norton and Schimel 2011) and subsequent production of N₂O through nitrification and denitrification processes (Toma and Hatano 2007). In all treatments, the cumulative N₂O and CO₂ productions during 4 weeks of incubation were linearly correlated with total mineral N concentration (NH_4^+ and

NO₃⁻) with $R^2 = 0.99$ and $R^2 = 0.98$, respectively. This implies that N2O and CO2 gas production was mainly related to N mineralization. Our observations agree with a previous report (Sánchez-Martín et al. 2008) where increasing N₂O production under aerobic condition was not correlated with the amount of dissolved organic matter but was related to the NH4+-N levels in the soil. Soils with different treatments showed different contributions to GWP. And seems that mostly GWP from the soils during the 4 weeks' incubation were produced when NH4⁺ content in soil was higher particularly in soil amended with urea. The total GWP from soils was within the range reported by Yu and Patrick (2004) at 8.6–250.6 kg CO₂ ha⁻¹ and Mu *et al.* (2013)'s range of 1.401–3.596 g CO_2 m⁻², while total GWP at urea amendment was highest. Therefore, there is much mitigation potential for N₂O production by using Azolla as a replacement for chemical fertilizer. During this experiment, the soil was at 55% of WHC, favoring nitrification over denitrification processes, and it is possible that the majority of N₂O was produced by nitrifiers. Nitrification is stimulated by high concentrations of NH4⁺ and does not require an organic C source (Norton and Stark 2011). The soil in this experiment had received long-term application of N fertilizers that may have resulted in increased potential for rapid nitrification. The significantly higher N2O production observed in soil treated with urea may also be explained by higher NH4⁺ leading to increased nitrification in ureatreated soil compared to those treated with Azolla compost, even with equivalent amounts of N applied (Tables 1 and 2). Therefore, a significant increase in cumulative N₂O production in urea versus Azolla treatments was observed even though both treatments had similar NO3contents. The higher NH4⁺ content in urea-treated soils throughout the incubation suggests these soils had favorable conditions for nitrification.

Nitrate N is the predominant form of N that is available in well-aerated soils, except for a short period after the addition of fertilizers containing NH_4^+ or urea (Stevenson and Cole 1999), and this experiment followed this generalization. Inorganic N pools were dominated by nitrate overall, with transient increases in the ammonium pool due to urea and its subsequent hydrolysis. Increases in the ammonium pool due to the Azolla high-level compost occurred during the first 3 weeks but were not evident by the fourth week. In contrast, the released ammonium from both compost and urea was nitrified and increased the nitrate pool during the incubation, although the timing of release was distinct for compost versus urea.

The mineralization processes of organic N and C are related to one another (Stevenson and Cole 1999). The mineralization of soil organic matter in Azolla compost treatments provides a continuous release of ammonium nitrogen (NH₄⁺-N) and nitrate nitrogen (NO₃⁻-N) (Tables 1 and 2). As expected, Azolla compost-amended soil released NH₄⁺ more slowly compared to the release of urea N to soils. Previous studies have also suggested that processes related to nitrification, including nitrifier denitrification, are a major source of N₂O emissions in fertilized and compost-amended soils (Zhu *et al.* 2013). Alternatively, increased ammonium also increases the availability of the nitrification product, nitrite or nitrate, that may be denitrified to N₂O in low-O₂ microsites in the soil.

Decomposition of the Azolla composts in the soil, as indicated by CO_2 production, was directly related to the amount of Azolla compost applied (Fig. 2). Azolla compost applications at the medium and high rates resulted in increased CO_2 production, reflecting readily available organic matter (Miyittah and Inubushi 2003). Increasing soil respiration rates would increase consumption of O_2 by microbes and may create microsites with the transiently anaerobic conditions required for denitrification. This is a probable explanation for our observation that N₂O production was significantly higher during the third week. This time period coincided with higher respiration combined with higher NO_3^- availability (Table 2), likely enhancing denitrification as well as nitrification sources of N₂O.

Sánchez-Martín et al. (2008) suggested that organic amendments instead of inorganic fertilizer may reduce the emission of N₂O and NO from agricultural fields. Rao et al. (1995) reported lower gaseous N losses from incorporated green manures than from urea applied in three splits in their field experiments, probably because mineralization of green manure N provided for more continuous N availability than urea application. Ding et al. (2013) reported that there was a significant difference between the nitrogen (N), phosphorus (P), and potassium (K) (NPK) fertilizers and organic matter and suggested that increasing Soil Organic Carbon (SOC) by compost application, then partially increasing N supply to crops versus adding inorganic N fertilizer alone, may be an effective measure to mitigate N₂O emissions from arable soils in the North China plain region.

Azolla compost treatments enhanced the height and dry weight of plants. Compost contains a proportion of available organic C and N that can increase soil fertility (Rao *et al.* 1995; Warman 2005; Diacono and Montemurro 2010; Vano *et al.* 2011) as well as inducing the activity of enzymes and microbial biomass soil (Zaman *et al.* 2002; Ros *et al.* 2006). Therefore, Azolla compost would stimulate soil microbes and increase nutrient recycling and decomposition in soil. Amir *et al.* (2012) also observed similar results in a similar pot experiment examining the growth of spinach (*Amaranthus tricolor* L.). They observed that Azolla compost treatments and urea fertilizer both increased the height and dry weight of spinach plants.

Our observations suggest that using Azolla compost as an amendment has the potential to reduce N_2O emissions when compared to urea fertilization. Azolla compost application may be particularly useful for vegetable crops in organic farming systems. In addition, Azolla treatments showed a reduction of atmospheric radiative forcing compared to urea-amended soils. Azolla compost is a viable amendment considering the agronomic benefits provided, while decreasing the likelihood of N_2O production under moderate moisture conditions. Further study is needed for long-term evaluation in organic farming systems as examined by Nagano *et al.* (2012).

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REFERENCES

- Amir L, Sari AP, Hiola SF, Jumadi O 2012: The availability of nitrogen in soil and growth of spinach. *Sainsmat. J.*, 1, 167–180. (in Indonesian).
- Azmal AKM, Marumoto T, Shindo H, Nishiyama M 1996: Mineralization and changes in microbial biomass in water-saturated soil amended with some tropical plant residues. Soil Sci. Plant Nutr., 42, 483–492. doi:10.1080/ 00380768.1996.10416317
- Azmal AKM, Marumoto T, Shindo H, Nishiyama M 1997: Changes in microbial biomass after continuous application of Azolla and rice straw in soil. *Soil Sci. Plant Nutr.*, 43, 811–818. doi:10.1080/00380768.1997.10414647
- Bordoloi LJ, Bhatt BP, Brajendra 2007: Effect of organic plant nutrient sources on groundnut (*Arachis hypogaea*) productivity and soil fertility under intensive integrated farming system in Meghalaya. *Environ. Ecol.*, 25, 1146–1150.
- Bouwman AF 1996: Direct emission of nitrous oxide from agricultural soils. Nutr. Cycl. Agroecosyst., 46, 53-70. doi:10.1007/BF00210224
- Bouyoucos CJ 1962: Hydrometer method improved for making particle size analyses of soils. *Agron. J.*, **54**, 464–465. doi:10.2134/agronj1962.00021962005400050028x

- Bray RH, Kurtz LT 1945: Determination of total organic and available forms of phosphorus in soils. Soil Sci., 59, 39–46. doi:10.1097/00010694-194501000-00006
- Burt R (ed.) 2004: Soil Survey Laboratory Methods Manual, Soil Survey Investigation Report N0.42 Ver. 4.0 USDA-NRSC, Lincoln, NE.
- Cheng W, Sakai H, Matsushima M, Yagi K, Hasegawa T 2010: Response of the floating aquatic fern Azolla filiculoides to elevated CO₂, temperature, and phosphorus levels. *Hydrobiologia*, 656, 5–14. doi:10.1007/s10750-010-0441-2
- Diacono M, Montemurro F 2010: Long-term effects of organic amendments on soil fertility. A review. Agron. Sustain. Dev., 30, 401–422. doi:10.1051/agro/2009040
- Ding WX, Luo JF, Li J, Yu HY, Fan JL, Liu DY 2013: Effect of long-term compost and inorganic fertilizer application on background N₂O and fertilizer-induced N₂O emissions from an intensively cultivated soil. *Sci. Total Environ.*, 465, 115–124. doi:10.1016/j.scitotenv.2012.11.020
- Djojosuwito S 2000: Azolla and Multipurpose Organic Farming. Jakarta, Kanisius Press (in Indonesian).
- Fukumoto Y, Inubushi K 2009: Effect of nitrite accumulation on nitrous oxide emission and total nitrogen loss during swine manure composting. *Soil Sci. Plant Nutr.*, 55, 428–434. doi:10.1111/j.1747-0765.2009.00376.x
- Herrero M, Thornton PK, Notenbaert AM *et al.* 2010: Smart Investments in Sustainable Food Production: revisiting Mixed Crop-Livestock Systems. *Science*, **327**, 822–825. doi:10.1126/science.1183725
- IBM Corp Released 2008: IBM SPSS Statistics for Windows, Version 16.0, IBM Corp, Armonk, NY.
- IPCC 2007: Climate Change 2007: Synthesis Report. IPCC, Geneva, Switzerland, 104 p.
- Jumadi O, Hala Y, Anas I, Ali A, Sakamoto K, Saigusa M, Yagi K, Inubushi K 2008a: Community structure of ammonia oxidizing bacteria and their potential to produce nitrous oxide and carbon dioxide in acid tea soils. *Geomicrobiol. J.*, 25, 381–389. doi:10.1080/01490450802402943
- Jumadi O, Hala Y, Inubushi K 2005: Production and emission of nitrous oxide and responsible microorganisms in upland acid soil in Indonesia. *Soil Sci. Plant Nutr.*, **51**, 693–696. doi:10.1111/j.1747-0765.2005.tb00093.x
- Jumadi O, Hala Y, Muis A, Ali A, Palennari M, Yagi K, Inubushi K 2008b: Influences of chemical fertilizers and a nitrification inhibitor on greenhouse gas fluxes in a corn (Zea mays L.) field in Indonesia. *Microbes Environ.*, 23, 29–34. doi:10.1264/jsme2.23.29
- Kimura SD, Yagi K, Mishima SI 2011: Carbon resources of residue and manure in Japanese farmland soils. Nutr. Cycl. Agroecosyst., 89, 291–302. doi:10.1007/s10705-010-9394-0
- Kirk TH, Beare MH, Meenken ED, Condron LM 2013: Soil organic matter and texture affect responses to dry/wet cycles: effects on carbon dioxide and nitrous oxide emissions. Soil Biol. Biochem., 57, 43–55. doi:10.1016/j. soilbio.2012.10.008
- Komilis D, Kontou I, Ntougias S 2011: A modified static respiration assay and its relationship with an enzymatic test to

assess compost stability and maturity. *Bioresource Tech.*, **102**, 5863–5872. doi:10.1016/j.biortech.2011.02.021

- Komilis D, Tziouvaras I 2009: A statistical analysis to assess the maturity and stability of six composts. Waste Manag., 29, 1504–1513. doi:10.1016/j.wasman.2008.10.016
- Kong YH, Nagano H, Kátai J, Vágó I, Oláh AZ, Yashima M, Inubushi K 2013: CO₂, N₂O and CH₄ production/consumption potentials of soils under different land-use types in central Japan and eastern Hungary. *Soil Sci. Plant Nutr.*, 59, 455–462. doi:10.1080/00380768.2013.775005
- Lang M, Cai Z, Chang SX 2011: Effects of land use type and incubation temperature on greenhouse gas emissions from Chinese and Canadian soils. J. Soils Sediment., 11, 15–24. doi:10.1007/s11368-010-0260-0
- Lou Y, Ren L, Li Z, Zhang T, Inubushi K 2007: Effect of rice residues on carbon dioxide and nitrous oxide emissions from a paddy soil of subtropical China. Water, Air, Soil Poll., 178, 157–168. doi:10.1007/s11270-006-9187-x
- Lumpkin TA, Plucknett D 1980: Azolla: botany, physiology, and use as a green manure. *Econ. Bot.*, 34, 111–153. doi:10.1007/BF02858627
- Miyittah M, Inubushi K 2003: Decomposition and CO₂-C evolution of okara, sewage sludge, cow and poultry manure composts in soils. *Soil Sci. Plant Nutr.*, **51**, 849–860.
- Mu ZJ, Huang AY, Ni JP, Li JQ, Liu YY, Shi S, Xie DT, Hatano R 2013: Soil greenhouse gas fluxes and net global warming potential from intensively cultivated vegetable fields in southwestern China. J. Soil Sci. Plant Nutr., 13, 566–578.
- Nagano H, Kato S, Ohkubo S, Inubushi K 2012: Emissions of carbon dioxide, methane, and nitrous oxide from shortand long-term organic farming Andosols in central Japan. *Soil Sci. Plant Nutr.*, 58, 793–801. doi:10.1080/ 00380768.2012.739550
- Norton JM, Schimel JP 2011: Nitrogen mineralization-immobilization turnover. *In* Handbook of Soil Science, Huang PM, Li Y and Summers ME, pp. 8–18. CRC Press, Boca Raton, FL.
- Norton JM, Stark JM 2011: Regulation and measurement of nitrification in terrestrial systems. *Methods Enzymol.*, 486, 343–368. doi:10.1016/B978-0-12-381294-0.00015-8
- Padre AT, Tsuchiya K, Inubushi K, Ladha JK 2005: Enhancing soil quality through residue management in a rice-wheat system in Fukuoka, Japan. Soil Sci. Plant Nutr., 51, 849–860. doi:10.1111/j.1747-0765.2005.tb00120.x
- Page AL, Miller RH, Keeney DR 1982: Methods of Soil Analysis, Part 2- Chemical and Microbiological Properties, 2nd Ed. American Society of Agronomy, Madison, WI.
- Rao KV, Kundu DK, Surekha K, Gandhi G 1995: Comparative efficiency of green manure and urea-N as affected by water deficit in lowland rice. IRRI-International Rice Research Institute. Fragile lives in fragile ecosystems, 13–17 February 1995, Eds. Los Banos, Philippines, pp. 256–266. Proceedings of the International Rice Research Conference, Philippines.
- Rayment GE, Higginson FR 1992: Australian Laboratory Handbook of Soil and Water Chemicals Methods. Australian Soil Land Survey Handbook, Inkata Press, Melbourne, Sydney.

- Ros M, Pascual JA, Garcia C, Hernandez MT, Insam H 2006: Hydrolase activities, microbial biomass and bacterial community in a soil after long-term amendment with different composts. *Soil Biol. Biochem.*, 38, 3443–3452. doi:10. 1016/j.soilbio.2006.05.017
- Rynk R. 1992. On-Farm Composting Handbook. Pub. 54. Northeast Regional Agricultural Engineering Service. Ithaca. NY.
- Sánchez-Martín L, Vallejo A, Dick J, Skiba UM 2008: The influence of soluble carbon and fertilizer nitrogen on nitric oxide and nitrous oxide emissions from two contrasting agricultural soils. *Soil Biol. Biochem.*, 40, 142–151. doi:10.1016/j.soilbio.2007.07.016
- Shibahara F, Inubushi K 1997: Effects of organic matter application on microbial biomass and available nutrients in various types of paddy soils. *Soil Sci. Plant Nutr.*, 43, 191–203. doi:10.1080/00380768.1997.10414727
- Singla A, Dubey SK, Iwasa H, Inubushi K 2013: Nitrous oxide flux from komatsuna (*Brassica rapa*) vegetated soil: a comparison between biogas digested liquid and chemical fertilizer. *Biol. Fertil. Soil*, **49**, 971–976. doi:10.1007/ s00374-013-0796-4
- Soil Survey Staff 1999: Soil Taxonomy: A Basic System of Soil Classification for Making and Interpreting Soil Surveys. 2nd Ed. Natural Resources Conservation Service. Handbook 436. Washington DC, USA, U.S. Department of Agriculture.
- Stevenson FJ, Cole MA 1999: Cycles of Soil Carbon, Nitrogen, Phosphorus, Sulfur, Micronutrients, 2nd ed., Wiley, New York.
- Talley SN, Talley BJ, Rains DW 1977: Nitrogen fixation by Azolla in rice fields. *In* Genetic Engineering for Nitrogen Fixation, Ed. Hollaender A, pp. 259–281. Plenum Press, New York and London.
- Thangarajan R, Bolan NS, Tian GL, Naidu R, Kunhikrishnan A 2013: Role of organic amendment application on greenhouse gas emission from soil. *Sci. Total Environ.*, 465, 72–96. doi:10.1016/j.scitotenv.2013.01.031
- Toma Y, Hatano R 2007: Effect of crop residue C:N ratio on N₂O emissions from Gray Lowland soil in Mikasa,

Hokkaido, Japan. Soil Sci. Plant Nutr., 53, 198–205. doi:10.1111/j.1747-0765.2007.00125.x

- Vano I, Matsushima M, Tang C, Inubushi K 2011: Effects of peat moss and sawdust compost applications on N₂O emission and N leaching in blueberry cultivating soils. *Soil Sci. Plant Nutr.*, 57, 348–360. doi:10.1080/ 00380768.2011.574596
- Velthof GL, Bruggen C, Groenestein CM, Haan BJ, Hoogeveen MW, Huijsmans JFM 2012: A model for inventory of ammonia emissions from agriculture in the Netherlands. *Atmospheric Environ.*, 46, 248–255. doi:10.1016/j. atmosenv.2011.09.075
- Warman PR 2005: Soil fertility, yield and nutrient contents of vegetable crops after 12 years of compost or fertilizer amendments. *Biol. Agricul. Horti.*, 23, 85–96. doi:10. 1080/01448765.2005.9755310
- Xu X, Inubushi K 2005: Mineralization of nitrogen and N₂O production potentials in acid forest soils under controlled aerobic conditions. *Soil Sci. Plant Nutr.*, **51**, 683–688. doi:10.1111/j.1747-0765.2005.tb00091.x
- Yu K, Patrick Jr WH 2004: Redox window with minimum global warming potential contribution from rice soils. *Soil Sci. Soc. Am. J.*, 68, 2086–2091. doi:10.2136/ sssaj2004.2086
- Zaman M, Cameron KC, Di HJ, Inubushi K 2002: Changes in mineral N, microbial biomass and enzyme activities in different soil depths after surface applications of dairy shed effluent and chemical fertilizer. *Nutr. Cyc. Agroecosys.*, 63, 275–290. doi:10.1023/A:1021167211955
- Zbytniewski R, Buszewski B 2005: Characterization of natural organic matter (NOM) derived from sewage sludge compost. Part 2: multivariate techniques in the study of compost maturation. *Bioresource Tech.*, 96, 479–484. doi:10.1016/j.biortech.2004.05.019
- Zhu X, Silva CRS, Doane TA, Wu N, Horwath RH 2013: Quantifying the effects of green waste compost application, water content and nitrogen fertilization on nitrous oxide emissions in 10 agricultural soils. J. Environ. Qual., 42, 912–918. doi:10.2134/ jeq2012.0445