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South Sulawvesi, m222 Indonesia Tel + 62-t5299974'l(AFa\ +62-411492646 E-mail rosdiana. gittutg@yahoo con Herry Sonjaya Depa(ment olAnimal Production. Fsculty ofAnimal tlusbndry, Hasanuddin University, Malassar' South Sula\$tsi, m245, Indoncsia Tcl: +62-4 l l 58428 l Fax +62-41 l 5872 l7 E-mail : ronj ayaheft)@gnoil.con Syamsuddin Hasan D€psrtment ofAnimal Nutrilion, Faculty ofAmmal Husbandry, Hasanuddin UnNersity, Makassar, South Sula\resi, 90245, Indonesia Tel: +62-81 1465771 Faxr +62-41 1587217 E-mail syon.hasah@tohoo.con Wempie lkpartm€nt ofAnimal Production.

Fsculty ofAnimal Husbadry, Hasanuddin University, Makassar, South Sulawtsi, 90245, Indoncsia Tel: +62-813439851 14 Fax +62411587211 E-mail pt)enpie@tshoo.con Abstrcttr Morica goat is an endemic species that can be found only in South Sulawesi, Indonesia. It has b€en reported by the FAO as enclargered species becaus€ its number of **population has been significantly** decreased.

Domestication by local people has been done by cultivating them with (acaltg goat that leads to an interbreeding process. Consenation of the goats must be done immediately due to its raprdly decreasing population. However, accurate identification and quanlification ofthe goat by DNA analysis is highly important.

The objective ofthis study is to determine the differences amongst marica- kacmg and Capra hircus. C. caucasicd and C- lalcohei from lhc G€nBank. The sequence of the mitochondrial DNA (mtDNA) at d-loop region of J0 Marica goats and five K4carrg goats from three districts in South Sula\$esi were investigated. Their nucleotide sequences were comparcd with the sequence ofthe CenBank's Capla sp afld were analyzed using Dendogram neighbor joining tr€e.

The results showed tha! There were a few nucleotide difrences between some Maica and Kacang goat that were located at 20,840 and 980bp In addition. both nucleotides sequence have shon genetic dislance compared to C ,i/crrj. However, comparing with other Caprd sp, the distance was significantly far. Meanwhile, according to the dendogram, it \ras found that all Goats and aapra sp came liom the sam€ ancestral

lineage.

It can be concluded that Marica and related goats could be very closely related with *C. capra*, but they were different from the *C. caucasica* and *C. dolgoprattensis*. Marica goat, Kacahg goat - *Capra hircus* South Sulawesi, mtDNA. Introduction One of Indonesian's germplasm commodities are goats. Goats spread in different regions with different climates and separated in a long time.

Various selections of environmental factors and treatment cause genetic changes in goat's population (Roul et al., 2008). Marica goat is a type of local goat that can be found only in South Sulawesi Province, especially in the district of Maros, Jeneponto, Soppeng and Makassar (Fitra et al., 2009). The goats are genetically potential to adapt in the dry land with a very low annual rainfall - International Journal of Agriculture Systems (IJAS) journal.

Manca Soats can survive in the dry season although they only eat dry grass in the rock ground. Unfortunately, according to Food and Agriculture Organization (2000) Report, its population is very low and is categorized as endangered species. Traditionally, local communities raise various kinds of goats in the same field and cages. It also happened on Marica Goat that was reared together with domestic goat.

consequently interbreeding and genetic mixing among Soat are inevitable. According to previous observation and interview with a local traditional breeder, Marica Soats have strong characteristic even they resulted from the combinations of Marica and Kacahg goats. Thus,

even the goats have strong characteristics of Marica goat, it does not guarantee that they are original Marica Goats. **Molecular genetic studies within and across breeds are essential for the** population management (Hall and Bradley, 1995, Ruane, 2000; Simianer, 2005). Performing such purpose, it is believed that using mitochondria DNA (mtDNA) is simpler than using nuclear DNA.

Mammalian mtDNA shows several special features such as absence of intron, maternal inheritance, lack of recombination events and a high mutation rate (Irwin et al., 1991). Furthermore, it brings enough information to figure out goat ancestries (Chen et al., 2005). Complete sequences of bovine mtDNA were published by Anderson et al.,

(1982), so the mtDNA have **widely used for genetic diversity and phylogenetic analysis among different cattle breeds** (Bradley et al., 1996; Troy et al., 2001). Specifically, D-loop area at mtDNA, non coding area, has nucleotide sequence that is **useful to determine**

phylogenetic relationship (Hou et al., 2006). Meanwhile, the preservation of Marica goats should be done immediately considering the number of its population has declined sharply.

However, before conducting conservation effort, determining the original Marica goat is essential. This study is therefore conducted to determine the kinship between Marica goat and Kacang goat and the original Marica from Kacang goats. 2. Materials and Methods 1.1 Animal Sample DNA was obtained from whole blood sample because the DNA has specific genetic characteristics and its size is smaller than nucleus DNA (Chen et al., 2005).

It was impossible to find a number of Marica goats that have the same characteristics due to the lack of its number. All Marica goats comprising of seven goats from Maros (T85, TB4, S1, S1, S2, T86 and TB8), three goats from Makassar (MK3, MK4 and MK5) and 20 goats from Jeneponto (JNP induk, JNP anak, MCJ2, MCJ3, I, Btg1, Btg2, Btg3, Btg4, TB VIII, B1, 82, 83, K1, K2, K1, Mks7, Mks8, Mks10, MksTJantan) were analysed. Almost all the goats were female except MksTJantan which was male.

On the other hand, five Kacang goats were taken from Makassar. 1.2 DNA isolation A total of 10 ml of blood were taken aseptically from the jugular vein of each Marica and Kacang goats and placed in vacuum glass tubes containing anticoagulant ACD (acid citrate dextrose). All samples were stored in a cooler and brought to laboratory for further analysis (Yadav and Yadav, 2008).

1.3 PCR Amplification In this study, only mitochondria DNA was used and purified following the method of Sambrook et al. (1989). 2.3 PCR Amplification The primers that were used in this study were designed to amplify D-loop region (Table 1). Primer for the D-loop was designed based on data *Capra hircus* sequence (GenBank Accession number: U295658) Program of Primer3 output (http://www-genome.wi.mit.edu/cgi-bin/primer3.cgi?results_from=primer3) was used to select primers which are predicted to give good results. PCR Reaction consisted of 2.5 mM MgCl₂,

10 mM dNTPs, 100-300 ng template DNA, 10 pmol each primer and 2U Taq Polymerase (Bio Lab) and its buffer. Table 1. Base sequences and its melting temperatures of primers for amplifying D-loop region of mtDNA of the goats. PRIMER Base sequence Number of Base Melting Temperature (°C) 59.0 The thermal profile included an initial denaturation at 94°C for 2 min followed by denaturation at 94°C for 2 minutes, annealing at 52°C for 2 minutes and 35 cycles of elongation and extension at 72°C for 1 min.

using PCR amplification. In figure PCR products were migrated at 1.270 agarose gel

run using 1xTBE buffer Submarine Electrophoresis [Hoefer, USA]. The gel was stained with ethidium bromide and observed under UV Transluminator (λ = 300nm) exposure, and the 1000 bp of standard molecular weight as a ladder. Purification of PCR product using Phylogenetic Analysis Products of PCR amplification was purified using GFX purification kit (Amersham, USA). Alignment of sequences was obtained using the Clustal W program (Thompson et al, 1994). The result was compared with DNA sequence of Capra hircus (GenBank Accession number GU295658.1), Capra hircus breed Inner Mongolia White (GenBank: Accession number GU068045.1), Capra hircus (GenBank Accession number: FJ207525.1) and Capra caucasica (GenBank Accession number: JN632609.1).

3. Results and Discussion

Figure 1 shows the profile of PCR amplification products of mtDNA gene. Some of the DNA bands show a smear and even no bands at all.

These might be due to the degradation process during DNA purification or the DNA chains were cut shorter during purification process. Some DNA do not form visible band that could be caused by the small amount of sample or analytical temperature. As a result, only 11 bands are clearly seen which are from B2F, r331 Target Name R/F D-loop CHF 5' CTCACATTA, AACCTGAGTC, C 3' 20 bp. (Journal of Agricultural Systems IUAS)

Figure 1 The results of PCR amplification of D-loop region of mtDNA of Marica and kacang goats (Ladder: 1000 bp). Table 2.

Sequences comparison of the nucleotides polymorphisms of D-loop DNA region

lc_Hr3 tB_2_F, BTG4-F, B1nr_F, a<_3_F, *Gcanll-F, \$TK-3.F, JfB_7_F, tTB_8-o-F, ts3_F, AATCCTACGA TCAATTCCCA ACAAACTAGG AGG\GTCCTA 401 401 401 401 401 401 401 401 401 401 401 c.. BTG4F, I-Betina-F, K3-F, JNP Induk_F, S3_F, Mk 3_I 1B_7_F and TB_8_o_F for Marica goats, kacang I F for kacang goat.

Table 2 illustrated some pertinent information about nucleotides sequence in the range 40bp. Almost all Marica and Kacang goat showed the same nucleotide sequences as C, i/c6 except for the parent JNP which is Marica goat sample from Jeneponto. This goat has nucleotide c, 1osine at 20bp while others have nucleotide adenine.

In addition, there are similar trend occurred in the range 960 and 1,000bp (Table 3) excluding TB_7_F and Kacang_I_F, respectively. Sample TB_F_7 which is Marica goat from Jeneponto has nucleotide cytosine while the other Marica, Kacang and C. hircus goats have thymine. On the other hand, at position 980bp, Kacang_I_F (Kacang goat) has guanine while other goats have adenine. Meanwhile, in the region of 840bp, two Marica goats (S3-F and MK_3F) show a difference where at 107bp, both of them have adenine nucleotide.

while the others have guanine Table 4 described the differences of nucleotide sequence between the researched Soats and the goat from CenBank which are *C. causica*, *C. falconeri* and *C. hitcus* inner Mongoli. white cashnirc. It can be clearly seen that the genetic distance have no difference between Marica Eoat utd Ir39 volume I lssLre 2 Decenroer 2013 E Table 3 Sequences comparison of the nucleotides polymorphisms of Dloop region of mtDNA, range E40 bp, 960 bp and 1,000 bp. #C_hircus dBj2,F #BTG_4-r #I_Bctm_F #K_3_F #JNP_hduk F *K...nLI_F #MK-3-F #TB-7-F #TB_E_o_F #s3 F 8401 8401 8401 8{01 E40l 8401 8401 E40l #C_hircus tB_2_F #BTG-4.F #I_Betn|a_F #K_3_F #JNP_induk_F #Ka.!nLI_F #MK-]-F #TB-7-F #TB_8_o_F #s3_F ATAAAGACAT AATATGTATATCGTACATTAACCATCICC [%O] 960l 960l 960l 960l 960l 960l 960l 960l e60l 960l 960l f€_hircts IB-?_F #BTG_4.F #I_B.tim_r #K-3_F #JNP_induk F #r'r.rnll_F }MK-3-F #TB-1-F #TB_t_o_F ,s3 f CCCATGCATA TAAGCACGTA CAATGTCCTTATTACCAGTA lIOOO} tr000l Ir000l Ir000l ltoool lI000l Ir0001 lImol tr 0001 l r0001 Ir000l C.

,rircrj Ttis trend is similar to KacarS goat and C /rirc,rj- Thrs condition can be explained by showing the dendogram Neighbor Joining Tree (Bootstrapping 1000 replications) as seen in Figure 2. It can be seen clearly that based on dendogram Neighbor Joining Tree, all Marica and Kacang goats have a far genetjc distance compare lo C /alconen and C.

catcsica, bvl their distance is closely related to species of C ,lircl/J. Meanwhile. a clos€r study on this dendogram shows that all goats sample has srmilar genelrc drslance. They are few subsritution mutalDns fotmd n Kacang ud Manca goats. Il might luppen as a resuli of adaptation proaess to environmenEl condition such as the lack of feed resources and selectron by breeders.

In Jeneponto, for cxample, the dry season duration is over seven months per year. Irol ATTTIATGAT CTACTTCACG TGTACGTACA TAATATTAAT C , 'G - nlernoliono Journolof Agriculllre SysteMs (UAS) Table 4. Matrix index ofnucleotide sequence's difference between Malica and Kacahg goats compare to the sequence liom the GenBank's goats.

t 2 3 4 5 6 7 t 9 l0 lt 12 13 14 2 B2F] BTG.4-F 00 000 5 K_I_F 0000 ttlt12 E MX-3-F trlrt22 9 TB_7_t tlllt222 l 53F 00000ltlt tttlt2202t 102 t02 102 r02 102 103 rot t03 t03 t02 t03 19 19 79 79 79 E0 E0 E0 80 79 m 106 l1 Cn||l*E 3E 3t 38 3t 18 39 39 19 39 IE l9 Et Figure 2.

Dendrogmm neighborjoining tree displaying the relationship between the sarnples and goats'panicular references b€longing to the *Capra* sp as determined by D.loop region of mtDNA gene analyses. The numbers at the node arc bootstrap values based on 1,000 re-samplings The bar represents the number of hutations per sequence position Thus, the feeding gass is less available and dry. Elrod dan Stansfield (2007) explained that the

total gene pool may change when phenotype characteristics is suitable with the environment.

Furthermore, the genes will be inherited to the offspring. Meanwhile, a research conducted by Agha et al (2008) concerning the mtDNA of Indian and European populations showed that the Indian and European breeds sampled from African and European populations seem to have differentiated from each other with only a little genetic exchange between the geographically isolated populations. Therefore, the mutation of population is often caused by genetic drift or natural selection (Nei and Kumar, 2000).

Conclusion It can be concluded that even though there are some differences in nucleotides in same Maica and Indian goats, all of the nucleotide sequences are similar to the nucleotide of C. (AF513441). However, it is different from C. caucasica and C. falconeri. Future work strongly recommended to perform the same analysis on the other area of D-loop mtDNA outside the area of 40,960 and 1,000bp to figure out other similarities and dissimilarities between Indian and Maica goats. Agha S H.F Pilla-S Caldi. M D'Andrea" S. Reale, A.Z.A. Abdelsalam and M H.

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